Preliminary Studies on the Secondary Metabolites of *Buchholzia Coriacea* (Wonderful Kola) Seed Ethanol Extract by GC-MS Analysis

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Abstract

Plants are the major sources of drugs. Natives all over the world use plants in different ways to treat diseases. Man, wild and domestic animals selectively eat these plants when they are sick. Our research interest is to find the phytochemicals in the seed of Bucholzia coriacea (Wonder cola) seed that produces its medicinal activity. Gas Chromatography-Mass Spectrometry (GC-MS) analysis was used for screening the secondary metabolites in its seed as a preliminary study. The extract was prepared using Soxhlet extraction method with ethanol as solvent. It was concentrated in a rotary evaporator. The molecular mass of the phytochemicals were established based on the fragmentation pattern in the mass spectra. The chromatogram showed twelve peaks representing the presence of twelve phytochemicals in the extract. The compounds were proposed based on comparison with National Institute of Standards and Technology (NIST) database. The suggested compounds are Beta-vinyl acrylic acid (10.62%), 2-methyl-pyrrolidine-2-carboxylic acid (33.60%), 1,2-benzenedicarboxylic acid, diethyl ester (0.89%), 14-ketopentadecanoic acid (0.68%), methyl 14-methylpenta decanoate (1.10%), norlinolenic acid (4.26%), hexadecanoic acid (13.26%), 9,12-octadecadienoic acid (5.64%), 9-octadecynoic acid (9.84%), linoelaidic acid (12.87%), micropine (1.02%) and anandamide (6.22%). The bioactivity studies showed that Bucholzia coriacea seed could be a urinary acidifier, inhibitor of uric acid production, catechol-O-Methyl transferase inhibitor which could control chronic infection like Parkinson's disease. We therefore recommend the isolation and total characterization of *B. coriacea* in order to confirm the secondary metabolites present.

Keywords: Bioactivity, extraction, gas- chromatography, Bucholzia coriacea, mass- spectrometry

INTRODUCTION

B. coriacea belongs to the family Capparaceae. It is commonly known as 'Wonderful kola' due to the medicinal properties of its seed and is widely seen in the rain forests of Nigeria, Cameroon, Liberia, Central African Republic, Congo, Ivory Coast and Gabon (Ezekiel and Onyeoziri, 2009; Mbata *et al.*, 2009). Traditionally it is known in Nigeria as 'uke' or 'Okpokolo' in Igbo, 'owi' in Edo and 'uworo' or 'Aponmu' in Yoruba (Ezekiel and Onyeoziri, 2009) while in other countries it is known as 'Ndo' in Mende (Sierra Leone), 'Doe-fiah' in Kru-basa (Liberia), 'Eson-bese' in Akanasante (Ghana), 'Banda' in Munga (West Cameroons), 'Essonbossi' in Central Africa and 'Kola Pimente' in French (Anowi *et al.*, 2012; Koudogbo *et al.*, 1972).

The plant is a medium sized evergreen tree growing up to 20m high with a smooth dark brown bark, wide leaves that are glossy and leathery arranged spirally and clustered with conspicuous cream white flowers in racemates at the end of the branches (Akpanyung et al., 1995; Culpeper, 1995). B. coriacea has a variety of medicinal uses which includes treatment of migraine when applied topically on the head (Erhirhie et al., 2015), anti-helminthic activity of the leaves and seed (Kameswararao et al., 2003) as well as antimicrobial properties (Nweze et al., 2006; Ejikeugwu et al., 2014). Obembe et al., 2012 results suggested that the extract of Buchholzia coriacea may have antifertility effects in male rats, the site of action most probably the epididymis . In Gabon B. coriacea is sometimes cultivated as both a medicinal and fetish plant (Lemmens, 2013). B. coriacea seed was shown to possess significant antidiabetic potential and also reduced lipid peroxidation in diabetic rats (Ezeigbo, 2011; Chinaka et al., 2012), while its leaf extract demonstrated antiinflammatory activity (Ezike et al., 2015). Our research therefore is aimed at studying the medicinal potentials and secondary metabolites in the ethanol extract of *B. coriacea* seed by GC-MS analysis. GCMS analyses has been used by several researchers to demonstrate the presence of primary and secondary metabolites in plant extracts (Igwe et al., 2016; Yan-qun et al., 2013; Igwe et al., 2016b; Divya and Subba, 2013; Igwe et al., 2016c). The pictorial view of **B. coriacea** is shown in Figure 1.



Fig -1: Pictorial view of B. coriacea

MATERIAL AND METHODS

a- Plant Materials

Fresh seeds of *B. coriacea* was harvested at Asaba in Delta State, Nigeria. The seeds were identified at the Taxonomy section of College of Natural,Resources and Environmental Management, Michael Okpara University of Agriculture, Umudike, Nigeria.

b- Preparation of Plant Extract

B. coriacea was dried in a shady place for 15 days and pulverized to powder using electrical grinder. Extraction was performed using soxhlet method (Jensen, 2007). Thirty five grams (36 g) of powdered sample was introduced into the extraction chamber of the soxhlet extractor using ethanol as solvent at a temperature of 70° C for 48 hrs. At the end of the extraction, the extract was concentrated in a rotary evaporator. The extract was sent for GC-MS analysis.

c- GC-MS analysis of B. coriacea

GC-MS QP2010 Plus (Shimadzu, Japan) was used in the characterization. The identification of the photochemical in the sample was carried out using a QP2010 gas chromatography with Thermal Desorption System, TD 20 coupled with mass spectrometry (Shimadzu). The ionization voltage was 70eV. Gas chromatography was conducted in the temperature programming mode with a Restek column (0.25 mm, 60m, XTI-5). The initial column temperature was 80°C for 1min, and then increased linearly at 70°C min-1 to 220°C, held for 3 min followed by linear increased temperature 10°C min-1 to 290°C for 10 min. The temperature of the injection port was 290°C and the GC-MS interface was maintained at 290°C. The sample was introduced via an all-glass injector working in the split mode, with helium carrier gas low rate of 1.2 ml min⁻¹.

d- Identification of secondary metabolites in B. coriacea

The GC-MS chromatogram of ethanol extract of *B. coriacea* was compared with the database of National Institute of Standards and Technology (NIST), NIST08.LIB (Stein, 1990), WILEY8.LIB (McLafferty, 1986) and with published literature. The name, molecular weight, formula, structure and bioactivities of the compounds were ascertained.

RESULTS AND DISCUSSION

B. coriacea gas chromatogram is presented in Figure 2. The mass spectra of *B. coriacea* seed is show in Figure 3.



Figure 2: Gas chromatogram of B. coriacea (wonderful kola) seed ethanol extract



Figure- 3: shows mass spectra of B. coriacea seed

S/No	Name of Compound	Retention	Peak	Molecular	Molecular	Bioactivity
,		time	Area (%)	weight	formula	
1	Beta-vinyl acrylic acid	3.389	10.62	98.09	C ₅ H ₆ O ₂	Antiamyloid-Beta, <u>Beta-</u> <u>2-Receptor-Agonist</u> , <u>Beta-Glucuronidase-</u> <u>Inhibitor</u>
2	2-Methyl-pyrrolidine-2- carboxylic acid	6.059	33.60	129.15	C ₆ H ₁₁ NO ₂	<u>Methyl-Guanidine-</u> <u>Inhibitor</u>
3	1,2-Benzenedicarboxylic acid, diethyl ester	11.952	0.89	222.23	$C_{12}H_{14}O_4$	Acidifier
4	14-ketopentadecanoic acid	15.449	0.68	256.38	$C_{15}H_{28}O_3$	<u>Acidulant</u>
5	Methyl 14- methylpentadecanoate	15.890	1.10	270.45	C ₁₇ H ₃₄ O ₂	<u>Catechol-O-</u> <u>Methyltransferase-</u> <u>Inhibitor</u>
6	Norlinolenic acid	16.317	4.26	264.40	C ₁₇ H ₂₈ O ₂	IncreaseAromaticAminoAcidDecarboxylase Activity
7	Hexadecanoic acid	16.544	13.26	256.42	C ₁₆ H ₃₂ O ₂	Arachidonic acid- Inhibitor
8	9,12-Octadecadienoic acid	18.251	5.64	294.47	$C_{19}H_{34}O_2$	Inhibit Production of Uric Acid
9	9-Octadecynoic acid	19.258	9.84	280.44	C ₁₈ H ₃₂ O ₂	Urine-Acidifier
10	Linoelaidic acid	19.406	12.87	280.44	C ₁₈ H ₃₂ O ₂	Acidifier

Table -1 Compounds and suggested bioactivity of B. coriacea (wonderful kola) seed ethanol extract

22.040

24.783

11

12

Micropine

Anandamide

H₂C

1.02

6.22

265.39

323.28

 $C_{16}H_{27}NO_2$

 $C_{20}H_{37}NO_2 \\$

Not found

Neurotransmitter

Figure- 4: Beta-Vinyl acrylic acid



Figure- 5: 2-Methyl-pyrrolidine-2-carboxylic acid



Figure- 6: 1,2-Benzenedicarboxylic acid, diethyl ester



Figure- 8: Methyl 14-methylpentadecanoate

Ο



Figure- 10: Hexadecanoic acid

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Figure 12: Anandamide

The GC-MS gas chromamatogram showed 12 peaks representing 12 phytochemicals in *B. coriacea* (wonderful kola) seed ethanol extract (Figure 2). The molecular weight is equivalent to the molecular ions in the mass spectra . The name, retention time, peak area percentage, molecular weight, molecular formular and bioactivity of B. coriacea (wonderful kola) seed ethanol extract are shown in Table 1 while their molecular structures are seen in Figures 4- 15. 2-Methylpyrrolidine-2-carboxylic acid with the highest peak area % of 33.60 and a retention time of 6.059 is a Methyl-Guanidine-Inhibitor (Duke, 1996). Methyl-Guanidine inhibitor, inhibits the action of the hydrolase enzyme methyguanidinase which is an <u>enzyme</u> that <u>catalyzes</u> the hydrolysis of methylguanidine to urea (Nakajima, 1980). Hexadecanoic acid at (RT: 16.544), (PA:13.26%) was the second abundant compound in the extract. It is an arachidonic acid inhibition. Beta-vinyl acrylic acid is an antiamyloid-beta-2-receptor agonist and beta-glucuronidase inhibitor. activity as analyzed by GC-MS. 9-Octadecynoic acid and Linoelaidic acid (RT: 19.258; 19.406) and (PA 9.84%; 12.87%) respectively are urine acidifier (Duke, 1996). Acidifiers are chemicals that reduce the pH of the body. Acidifiers are needed for food digestion especially in patients suffering from achlorhydria. These patients are not able to secret HCl for food digestion. These phytocompounds will be beneficial since it increases gastric acid when ingested. The compound 9,12-Octadecadienoic acid with peak area % 5.64 may inhibit uric acid production. Uric acid inhibitor is a compound that inhibits the acidification of urine minimizing the risk of formation of uric acid stones and deposition of uric acid crystals in the joints such as the toe and knee joints that form gout thereby reducing episodes of sharp pain in the affected joints (Muhammad, 2013). Methyl 14-methylpentadecanoate is a known <u>catechol-O-methyl transferase inhibitor</u>. A chronic infection like Parkinson's disease is treatable with a catechol-O-methyl-transferase inhibitors (Burkhard et al., 2001). Since methyl 14-methylpentadecanoate and 9,12-octadecadienoic acid, methyl ester are inhibitors of catechol-O-methyl-transferase (Duke, 1996), they may be effective in the treatment of Parkinson's disease. Catechol-O-methyltransferase is involved in the degradation of neurotransmitters but the inhibitors oppose the degradation of neurotransmitters. COMT is involved in the degradation of catecholamine (dopamine, noradrenaline and adrenaline) which are neurotransmitters. Anandamide, another compound with lower concentration in the extract (PA: 6.22%) and at (RT: 24.783) is responsible for neurotransmission action.

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REFERENCE

Akpanyung EO, Udoh, AP, Akpan EJ (1995): Chemical composition of the edible leaves of *Pterocarpus mildbreadii.* Plant Foods for Human Nutrition. 4 (3): 209.

Anowi FC, Ike C, Ezeokafor E, Ebere C (2012): The Phytochemical, Antispamodic and Antidiarrhoea properties of the methanol extract of the leaves of *Buchholzia coriacea family Capparaceae.* International Journal of Current Pharmaceutical Research. 4(3): 52-55.

Burkhard P, Dominici P, Borri-Voltattorni C, Jansonius JN, Malashkevich VN (2001): "Structural insight into Parkinson's disease treatment from drug-inhibited DOPA decarboxylase".

Nature Structural Biology. 8(11): 963 -967.

Chinaka ON, Okwoche JO, Florence CN and Nkeiruka EU (2012): Effects of Methanol Extract of *Buchholzia coriacea* Fruit in Streptozotocin-induced Diabetic Rats. Journal of Pharmacology and Toxicology, *7: 181-191.*

Culpeper NC. (1995): Complete Herbal: A Book of Remedies of Ancient Ills. The Word's Worth Reference Collection Library, Contemporary Publishing Company.

Divya NP, Subban R (2013): GC-MS analysis of essential oil obtained from *Heracleum candolleanum* (Wight et Arn). Elsevier: Journal of Pharmacy Research 6: 155 -157

Duke's Phytochemical and Ethnobotanical Databases (1996): U.S. Department of Agriculture, Agricultural Research Service. [Online]. Available: <u>http://phytochem.nal.usda.gov</u>

Ejikeugwu C, Umeokoli B, Iroha I, Ugwu M, Esimone C (2014): Phytochemical and Antibacterial Screening of Crude Extracts from Leaves of Wonderful Kola, American Journal of Life Sciences. Special Issue: Microbiology Research. 2(3-6): 9 -12.

Erhirhie EO, Ben-AzuBenneth, MG, Emuesiri CP and Omonjiahio IA (2015): Ethnopharmacological Review of *Buchhoizia coriacea* (Wonderful Kola).International Journal of Advances In Pharmacy, Biology And Chemistry 4(1):149 -155

Ezeigbo II (2011): The Antidiabetic Potentials of the Methanolic Seed Extract of Buchhlozia coriacea.Annals of Medical and Health Sciences Research, 1(2): 159–163.Ezekiel

OO, Onyeoziri NF (2009): Preliminary studies on the antimicrobial properties of *Buchholzia coriacea* (wonderful kola). African Journal of Biotechnology. 8(3): 472-474.

Ezike AC, Onyeto CA, Nwabunike IA, Mbaoji FN, Attah BE, Amanambu SO, Okoli CO (2015): Anti-Inflammatory Activity Of *Buchholzia coriacea* Engl. (Capparaceae) Leaf Extract: Evaluation Of Components Of The Inflammatory Response Involved. J Complement Integr Med. 2:153-158.

Igwe KK., Nwankwo PO, Otuokere IE, Chika I, Amaku FJ (2016): Studies on the medicinal plant *Acalypha Wilkesiana* ethanol extracts phytocomponents by GC-MS analysis, Global Journal of Science Frontier Research, 16(2): 48–55.

Igwe KK, Madubuike AJ, Otuokere IE, Amaku FJ, Chika I (2016): GC-MS analysis for structural identification and bioactive compounds in methanolic leaf extract of *Mallotus oppositifolius* International Journal of Scientific Research and Management, 4(5): 4123–4128

Igwe K, Madubuike AJ, Chika I, Otuokere IE, Amaku FJ (2016): Studies on the medicinal plant *Pausinystalia yohimbe* ethanol leaf extracts phytocomponents by GC-MS analysis, International Journal of Scientific Research and Management, 4(5): 4116–4122

Jensen WB (2007): The origin of Soxhlex Extraction. Journal Clinical Education. 84 (12): 1913 – 1914 Kameswararao B, Kesavulu MM, Apparao C (2003): Evaluation of antidiabetic effect of *Momordica cymbalaria* fruit in alloxan-diabetic rats. Fitoterapia. 74: 7 – 13.

Koudogbo B, Delaveau P, Adjanohoun E (1972): Study of an African Cepparidaceae, *Buchholzia Coriacea*, Engler. Ann Pharm Fr. 30 (2): 93-98

Lemmens, RHMJ (2013): Buchholzia coriacea Engl. In: Schmelzer GH and Gurib-Fakim A.(Editors). PROTA (Plant Resources of Tropical Africa / Ressources végétales de l'Afrique tropicale),Wageningen, Netherlands. 2013. Accessed 3 febuary 2015.

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McLafferty FW (1986): Registry of mass spectral data. Fourth electronic ed. Wiley New York Muhammad BH (2013): Reducing uric acid. International Centre for Diarrhoeal Research, Bangla. Research Gate.

Mbata TI, Duru CM, Onwumelu HA (2009): Antibacterial activity of crude seed extracts of *Buchholzia coriacea* E. on some pathogenic bacteria, Journal of Developmental Biology and Tissue Engineering. 1(1): 1 -5.

Nweze NE, Asuzu IU (2006): The antihelmintic effects of *Buchhlozia coriacea* seed. Nig Vet J., 27: 60 – 65.

Nakajima M, Shirokane Y, Mizusawa K (1980): A new amidino hydrolase, methylguanidineamidino hydrolase from Alcaligenes sp. N-42". FEBS Lett. **110** (1): 43–46.

Obembe OO, Onasanwo, SA, Raji Y (2012): Preliminary study on the effects of *Buchholzia Coriacea* seed extract on male reproductive parameters in rats. Niger. J. Physiol. Sci. 27: 165 – 169 **Stein SE (1990).** National Institute of Standards and Technology (NIST) Mass Spectral Database and Software Version 3.02,USA

Yan-qun Li, De-xin K, Hong W. (2013): Analysis and evaluation of essential oil components of cinnamon barks using GC-MS and FTIR spectroscopy. Elsevier: Industrial Crops And Products. 41: 269 – 278